

Mini Review

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Where are Cell-Based Therapies Heading? Current Limitations and Future Directions

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ABSTRACT

Congestive heart failure (CHF) secondary to chronic coronary artery disease is a major cause of morbidity and mortality worldwide. Its prevalence is increasing despite advances in medical and device therapies. Adult stem cell therapies have emerged as a promising treatment generating new cardiomyocytes and vessels, and are anticipated to reverse functional deterioration in patients with congestive heart failure for whom heart transplantation is the only cure. This field was enthusiastically studied in last two decades, revealing that the major beneficial outcomes from cell therapy are associated with paracrine effects rather than direct differentiation. Accordingly, paracrine factors (e.g., growth factors, cytokines and microRNAs) secreted from stem cells reduce scar volume and myocyte apoptosis, increase myocyte proliferation and activate endogenous cardiac stem cells to produce new myocytes. Moreover, functional efficacy of progenitor cells isolated from the bone marrow, adipose tissue and the heart have shown promising effects in preclinical animal models. These convincing results led to the initiation of clinical trials using autologous and allogeneic stem cells, and progenitor cells. Although clinical trials demonstrated their safety in humans, therapeutic efficacy is still inconclusive. This review discusses the progress and limitations of cell-based therapies and alternative solutions for future advancement.

KEY WORDS: Congestive heart failure (CHF); Myocardial Infarction (MI); Adult stem cells; Mesenchymal stem cells (MSCs); Cardiac stem cells (CSCs); Cardiosphere-derived cells (CDCs).

INTRODUCTION

Congestive heart failure (CHF) secondary to chronic coronary artery disease is a major cause of morbidity and mortality worldwide.¹ Its prevalence is increasing despite advances in medical and device therapies. Currently available medical interventions attenuate neurohormonal activation (e.g., renin-angiotensin-aldosterone system, sympathetic nervous system, and arginine vasopressin), reducing myocyte apoptotic cell death and interstitial connective tissue proliferation, and attenuating the progression of myocyte cellular hypertrophy. However, none of the current therapies are effective in reversing myocyte loss and cellular abnormalities associated with poor myocyte contractile performance which are impaired in the failing heart. Therefore, cardiac transplantation has been the only available cure for people who develop advanced heart failure.

Cell based therapy emerged as an alternative new therapy to restore impaired cardiac function.^{2,3} Over the past two decades, cell-based studies have been studied enthusiastically. Experimental studies demonstrated promising effects on generating new cardiomyocytes and vessels, reversing functional deterioration and preventing the progression to CHF.⁴ Furthermore, *in vivo* experiments revealed that the major beneficial outcomes from cell therapy are associated with paracrine effects, rather than direct regeneration of new tissue.^{5,6} Stem cells secrete paracrine factors (e.g., growth factors, cytokines and microRNAs), which reduce scar volume and myocyte apoptosis, increase myocyte proliferation and activate endogenous car-

diac stem cells to produce new myocytes.⁷⁻¹¹ Convincing data from preclinical animal models led to several clinical trials using autologous and allogeneic stem cells and progenitor cells to assess their safety in humans.¹²⁻¹⁴ However, their clinical relevance is still inconclusive.^{9,13} Accordingly, the therapeutic benefits of the majority of clinical studies are modest at most.¹⁴ The discrepancies between the animal studies and multiple clinical studies require reassessment of current strategies of cell-therapies. In the following sections we discuss current therapeutic limitations and alternative solutions (also summarized in Table 1) assessed in the experimental and clinical fields.

CURRENT LIMITATIONS AND ALTERNATIVE SOLUTIONS

Low Cell Retention Associated with Cell Delivery Approach

For now, three major cell delivery approaches have been tried in clinical applications (intravenous, intramyocardial (epicardial or subendocardial) and intracoronary injections). For intravenous injection, cells were injected systemically but only 0.04% of cells reached the heart and the majority of cells were entrapped in other organs (i.e., lung, kidney, liver and spleen).¹⁵ Intramyocardial and intracoronary injection approaches have relatively better outcomes on cell retention. However, within minutes of intramyocardial or intracoronary stem cell injection the majority of cells (~85% of cells) are washed out through the coronary venous system or mechanically ejected from the injection site and only 1-2% of the cells are retained in the heart 1-month post-injection.¹⁶⁻¹⁸ Therefore, to maximize their regenerative effects on the myocardium several approaches are being considered to enhance cell viability, improve functional properties of individ-

ual stem cells, and to prolong cell retention.

Enhancement of Cell Viability and Functional Properties with Genetic Modification

Genetic modification has been mostly assessed using bone marrow derived mesenchymal stem cells (MSCs). Since MSCs lack the expression of major histocompatibility complex (MHC) class II antigen, allogeneic MSCs can escape direct recognition of helper T-cells and are deemed immunoprivileged. The safety and efficacy of MSCs has been demonstrated by clinical work and there is increasing interest in enhancing the benefits of MSC therapy. For example, combining MSC and pharmacotherapy,¹⁹ genetically modifying MSCs²⁰⁻²² and pre-conditioning MSCs^{23,24} are approaches that are being explored to augment MSC-mediated cardiac repair. MSCs transfected to overexpress Akt or cell survival protein promote myocardial protective function.^{6,21} Furthermore, MSCs engineered to express combinations of gene products such as Akt and angiopoietin-1 (Ang1) have also shown functional benefits in experimental animal models.²⁵ MSCs overexpressing vascular endothelial growth factor (VEGF) and Stromal cell-derived factor-1 (SDF-1) improve cardiac function by activating the Akt pathway.²⁰ MSCs transfected to express heme-oxygenase 1 (HO-1), an enzyme that improves MSC tolerance to hypoxia, infused into a cardiac ischemia-reperfusion model improve EF and lower end systolic volume compared to controls.²⁶ Although, these preconditioned MSCs improve engraftment and survival of transplanted cells, due to safety concerns of genetic modification on stem cell nucleus, clinical application is unwarranted.

Table 1: Alternative Approaches to Overcome Current Limitation of Cell-based Therapy.

<u>Enhancement of cell survival, Mobilization and paracrine secretion</u>
- Pharmacology (Statins, etc.)
- Genetic modification (Akt and Ang1, VEGF and SDF-1, HO-1)
- Non-genetic modification (Hypoxia, bFGF/IGF-1/BMP2, poly(I:C), microRNAs)
<u>Enhancement of cell retention/survival</u>
- Biomaterials (hyaluronic acid, collagen, fibrin, ECM, peptide, polymer)
- Cell patch (Cell sheet, scaffold or scaffold-free bioprinting)
<u>Synergistic or accumulating effects of cell</u>
- Synergistic effects: MSCs and CSCs
- Accumulating effects: Repeated cell infusion (MSCs, CSCs, CDCs)

MSCs: Mesenchymal Stem Cells; CSCs: Cardiac Stem Cells; CDCs: Cardiosphere-derived cells; Ang1: Angiopoietin 1; VEGF: Vascular endothelial growth factor; SDF: Stromal cell-derived factor; HO-1: Hemeoxygenase-1; bFGF: Basic fibroblast growth factor; IGF: Insulin-like growth factor; BMP2: Bone Morphogenetic protein 2; Polyinosinic polycytidylic acid; ECM: Extracellular matrix.

Enhancement of Cell Viability and Functional Properties without Genetic Modification

Because genetically engineered stem cells may have unwanted long-term side-effects, pre-treatment of stem cells without genetic modification are considered more practical and relevant approaches. One method includes hypoxia preconditioning. Since cells are exposed to a harsh hypoxic environment after injection into the ischemic area, preconditioning of cells in a hypoxic chamber (1-3% O₂) prior to transplantation is reasonable and, in fact, can improve cell survival in this environmental stress scenario.²⁷ Hypoxia stimulation activates pro-survival pathways *via* phosphorylation of Akt and p38 resulting in HIF-1 α activation.^{23,28} Hypoxic preconditioning of MSCs²⁰ or cardiac stem cells (CSCs)²⁹ enhanced therapeutic effects in an ischemia model.

Another approach is pretreatment with growth factors. MSCs pretreated with Basic fibroblast growth factor (bFGF), insulin-like growth factor (IGF)-1 and bone morphogenetic protein 2 (BMP2) improved myocardial repair in a rat model of myocardial infarction (MI).³⁰ Behfar and Terzic et al pretreated MSCs with growth factors to enhance their cardioprotective functions. They demonstrated the ability of a “cardiogenic cocktail” (consisting of TGF β 1, BMP-4, Activin-A, retinoic acid, IGF-1, FGF-2, α -thrombin and IL-6) to enhance the therapeutic benefits of autologous MSCs. Subsequently the same group initiated clinical trials in patients with class 2 or 3 heart failure (C-CURE) trial.³¹ Also, the Safety and Efficacy of Autologous Cardiopoietic Cells for Treatment of Ischemic Heart Failure (CHART-1) trial is currently ongoing.

We demonstrated that stimulation of Toll-Like Receptor 3 (TLR3) produced many trophic factors without induction of inflammatory-related cytokines.²⁶ Poly (I:C) is structurally similar to double-stranded RNA and is known to interact with TLR3, which is expressed on the membrane of B-cells, macrophages, dendritic cells, bone marrow and heart-derived stem cells (MSCs and CDCs). Poly (I:C) directly reacts with the TLR3 receptor on the endosome of MSCs/CDCs. After stimulation with poly (I:C) MSCs/CDCs are collected and washed and since the poly (I:C) does not reside within the cells, it does not affect the heart environment after injection of cells. Interaction of poly (I:C) with TLR3 on MSCs causes secretion of the growth factor VEGF and the cytokine IL-6 without upregulation of the inflammatory cytokines IL-1 and tumour necrosis factor- α (TNF α). Injection of TLR3 activated MSCs (TLR3-MSCs) in a non-ischemic cardiomyopathy model improved cardiac function more than standard MSCs along with increased myocyte proliferation, reduced fibrosis and reduced myocyte apoptosis.³² Also activation of TLR3 on Cardiosphere-derived cells (CDCs) (TLR3-CDCs) stimulated the secretion of HGF, IGF1 and IL-6 without up-regulation of inflammatory cytokines.³³ Thus, TLR3-MSCs or TLR3-CDCs are safe and feasible to use in the human heart. Further investigation is necessary to confirm long-term

safety and feasibility in a preclinical animal model. Transient genetic modulation of cellular therapies may minimize unwanted side effects in the heart environment and can be considered clinically relevant approaches.

Recently, it was reported that exosomes secreted from stem cells play important roles for cardiac regeneration.^{8,11,30} Exosomes transfer microRNAs from cell-to-cell and inhibit inflammation⁸ (miR146a, miR155) and apoptosis³⁴ (miR21, miR22, miR24), and increase angiogenesis (miR210) and myocyte proliferation³⁵ (miR1, miR133a, miR294). Therefore, new methods of treatment are focusing on modulating microRNAs in stem cells. For example, Hu et al showed that a cocktail of three miRNAs (miR21, miR24, and miR22) was able to substantially improve the engraftment of CSCs by targeting apoptotic activators.³⁴ Similarly, in MSCs, miR133 was shown to act as a pro-survival agent to mitigate ischemic insult on the cells and improve subsequent engraftment.³⁵

Enhancement of Cell Retention by Bio-Injectable Materials and Cell Patch

The use of biomaterials is an effective approach to enhance cell retention and survival after implantation into damaged myocardium. Biomaterials physically support the cells to improve retention directly after administration and create a protective environment for the survival of the cell. Mainly bio-injectable materials are injected into myocardium or cell patch is placed on the surface of myocardium.

Recently, injectable biomaterials combined with cell-based therapies for cardiovascular disease are gaining more attention because they have shown therapeutic potential in preclinical models for MI. Natural (e.g., hyaluronic acid,³⁶ collagen^{37,38}, fibrin³⁹⁻⁴¹ or extracellular matrix-based⁴²) or synthetic (e.g., peptide⁴³ or polymer-based⁴⁴) materials can enhance stem cell survival and retention *in vivo*, prolong growth factor release from hydrogel or particle constructs and stimulate endogenous cardiac regeneration. Although, there is promising preclinical data, the therapeutic potential of biomaterial-based products for cardiovascular disease has yet to be proven in a clinical setting.

Cell Patch technology generates a tissue-like structure *in vitro* and transplants it, typically onto epicardial myocardium. The main advantage of this approach is that the cells are cultivated under precise culture conditions. Therefore, cell proliferation, differentiation, and tissue structure can be well-controlled. Nevertheless, there are several limitations of this approach *in vivo*. 1) This procedure is more invasive than catheter-based approaches since open chest surgery is required. 2) Poor nutrient diffusion and vascularization immediately after transplantation usually limits the thickness of the constructs and long-term cell survival in the heart.¹⁶ 3) Patch-based transplantation provides inadequate integration of the graft with the host myocardium. Although paracrine factors secreted from cell patches can eas-

ily cross the barrier and be effective, poor vascularization and improper coupling of cultured cardiomyocytes with the native myocytes may limit remuscularization and therapeutic efficacy. Recently, 3D bioprinting technology has been introduced in the cardiovascular field.⁴⁵ This uses 3D structured cell spheroids rather than a monolayer of cells since 3D structure is known to enhance hypoxia resistance and encourage vascularization.^{46,47} The application of this technology may overcome current limitations associated with patch-based therapy.

Combination of Mesenchymal Stem Cells and Cardiac Stem Cells

Another approach is combined MSC and CSC to enhance the therapeutic effects of each cell type. Recent work by Williams et al demonstrated that the combined use of 1 million human CSCs with 200 million human MSCs provided greater recovery, almost to baseline, in a swine model of anterior wall MI.⁴⁸ While all stem cell treated animals demonstrated improved left ventricular ejection fraction compared to placebo controls, notably, animals receiving dual cell therapy had a 2-fold greater reduction in scar size (21.1% for CSC/MSC *versus* 10.4% for CSC alone or 9.9% for MSC alone) and had improved rates of pressure change during diastole. Overall left ventricular chamber dynamics were improved in both the dual therapy and CSC or MSC alone treated groups. Interestingly, CSC alone treated animals demonstrated better isovolumic relaxation as compared to controls, while MSC alone treated animals exhibited improved diastolic compliance, indicating that the enhanced effect of dual therapy on both systolic and diastolic function may be due to a synergistic effect between CSC and MSC targeted mechanisms. A current clinical trial has been initiated to assess the therapeutic effects (CONCERT-HF: ClinicalTrials.gov #NCT02501811).

Repeated Stem Cell Injection

Allogeneic MSCs and CDCs are immunoprivileged and can escape from direct recognition of helper T-cells due to the lack of expression of MHC class II antigen.^{49,50} Based on these observations, a recent clinical trial was initiated using allogeneic human MSC/CDC treatment in patients with chronic myocardial infarction (POSEIDON⁵¹, ALLSTAR⁵⁹). Since a single injection of MSCs or CDCs has moderate influence on cardiac function and reduced scar volume^{52,53}, it was thought that repeated injections of stem cells would be more effective in regenerating myocardial tissue.^{54,55} However, the initial infusion of cells activates and enhances the immune response^{49,50} and subsequent injected cells are quickly eliminated and ineffective. This quick reaction is mainly associated with acquired/adaptive rather than innate immunity. Thus, development of efficacious MSC/CDC platforms administered with optimal immune suppression could circumvent barriers related to multiple injections of stem cells and allow the widespread application of “off-the-shelf” cell therapy to treat the large number of patients in need.^{50,56,57} Gene-editing technology could also be applied to minimize acquired immunity creating immune-tolerant MSCs or CDCs.

CONCLUSION

Promising data derived from experimental models indicate the potential success of using cell based therapy in clinical applications. However, early stage clinical trials are revealing therapeutic limitations. We need to reassess the current problems and find alternative solutions. Feedback from clinical outcomes is providing more information for the development of the second stage of cell-based therapy research. In light of their proven safety profiles, adult stem cells (i.e., bone marrow mononuclear cells, adipose-derived stem cells, MSCs, CDCs and CSCs) are prime candidates for cell based therapies. Genetic modification, preconditioning, biomaterials, bioengineering, combination of cells and repeated injection approaches will further improve the efficacy of stem cell therapy. Taken together, the current understanding of stem cell based therapy and the emerging approaches and discoveries will definitely advance cell-based therapy and cure many CHF patients.

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CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

REFERENCES

1. Mozaffarian D, Benjamin EJ, Go AS, et al; American Heart Association Statistics C, Stroke Statistics S. Heart disease and stroke statistics--2015 update: A report from the American Heart Association. *Circulation*. 2015; 131: e29-322. doi: [10.1161/CIR.0000000000000152](https://doi.org/10.1161/CIR.0000000000000152)
2. Suzuki G. Translational research of adult stem cell therapy. *World J Cardiol*. 2015; 7: 707-718. doi: [10.4330/wjc.v7.i11.707](https://doi.org/10.4330/wjc.v7.i11.707)
3. Anversa P, Kajstura J, Rota M, Leri A. Regenerating new heart with stem cells. *J Clin Invest*. 2013; 123: 62-70. doi: [10.1172/JCI63068](https://doi.org/10.1172/JCI63068)
4. Braunwald E. The war against heart failure: The lancet lecture. *Lancet*. 2015; 385: 812-824. doi: [10.1016/S0140-6736\(14\)61889-4](https://doi.org/10.1016/S0140-6736(14)61889-4)
5. Malliaras K, Zhang Y, Seinfeld J, et al. Cardiomyocyte proliferation and progenitor cell recruitment underlie therapeutic regeneration after myocardial infarction in the adult mouse heart. *EMBO Mol Med*. 2013; 5: 191-209. doi: [10.1002/emmm.201201737](https://doi.org/10.1002/emmm.201201737)
6. Mangi AA, Noiseux N, Kong D, et al. Mesenchymal stem cells modified with akt prevent remodeling and restore performance of infarcted hearts. *Nat Med*. 2003; 9: 1195-1201.

7. Hodgkinson CP, Bareja A, Gomez JA, Dzau VJ. Emerging concepts in paracrine mechanisms in regenerative cardiovascular medicine and biology. *Circ Res*. 2016; 118: 95-107. doi: [10.1161/CIRCRESAHA.115.305373](https://doi.org/10.1161/CIRCRESAHA.115.305373)
8. Ibrahim AG, Cheng K, Marban E. Exosomes as critical agents of cardiac regeneration triggered by cell therapy. *Stem Cell Reports*. 2014; 2: 606-619. doi: [10.1016/j.stemcr.2014.04.006](https://doi.org/10.1016/j.stemcr.2014.04.006)
9. Bolli R, Ghafghazi S. Cell therapy needs rigorous translational studies in large animal models. *J Am Coll Cardiol*. 2015; 66: 2000-2004. doi: [10.1016/j.jacc.2015.09.002](https://doi.org/10.1016/j.jacc.2015.09.002)
10. Orenes-Pinero E, Montoro-Garcia S, Patel JV, Valdes M, Marin F, Lip GY. Role of microRNAs in cardiac remodeling: New insights and future perspectives. *Int J Cardiol*. 2013; 167: 1651-1659. doi: [10.1016/j.ijcard.2012.09.120](https://doi.org/10.1016/j.ijcard.2012.09.120)
11. Sahoo S, Losordo DW. Exosomes and cardiac repair after myocardial infarction. *Circ Res*. 2014; 114: 333-344. doi: [10.1161/CIRCRESAHA.114.300639](https://doi.org/10.1161/CIRCRESAHA.114.300639)
12. Schachinger V, Assmus B, Britten MB, et al. Transplantation of progenitor cells and regeneration enhancement in acute myocardial infarction: Final one-year results of the topcare-ami trial. *J Am Coll Cardiol*. 2004; 44: 1690-1699. doi: [10.1016/j.jacc.2004.08.014](https://doi.org/10.1016/j.jacc.2004.08.014)
13. Gyongyosi M, Wojakowski W, Lemarchand P, et al. Meta-analysis of cell-based cardiac studies (accrue) in patients with acute myocardial infarction based on individual patient data. *Circ Res*. 2015; 116: 1346-1360. doi: [10.1161/CIRCRESAHA.116.304346](https://doi.org/10.1161/CIRCRESAHA.116.304346)
14. Abdel-Latif A, Bolli R, Tleyjeh IM, et al. Adult bone marrow-derived cells for cardiac repair: A systematic review and meta-analysis. *Arch Intern Med*. 2007; 167: 989-997. doi: [10.1001/archinte.167.10.989](https://doi.org/10.1001/archinte.167.10.989)
15. Freyman T, Polin G, Osman H, et al. A quantitative, randomized study evaluating three methods of mesenchymal stem cell delivery following myocardial infarction. *Eur Heart J*. 2006; 27: 1114-1122. doi: [10.1093/eurheartj/ehi818](https://doi.org/10.1093/eurheartj/ehi818)
16. Narita T, Shintani Y, Ikebe C, et al. The use of scaffold-free cell sheet technique to refine mesenchymal stromal cell-based therapy for heart failure. *Mol Ther*. 2013; 21: 860-867. doi: [10.1038/mt.2013.9](https://doi.org/10.1038/mt.2013.9)
17. Willmann JK, Paulmurugan R, Rodriguez-Porcel M, et al. Imaging gene expression in human mesenchymal stem cells: From small to large animals. *Radiology*. 2009; 252: 117-127. doi: [10.1148/radiol.2513081616](https://doi.org/10.1148/radiol.2513081616)
18. Nguyen PK, Lan F, Wang Y, Wu JC. Imaging: Guiding the clinical translation of cardiac stem cell therapy. *Circ Res*. 2011; 109: 962-979. doi: [10.1161/CIRCRESAHA.111.242909](https://doi.org/10.1161/CIRCRESAHA.111.242909)
19. Yang YJ, Qian HY, Huang J, et al. Combined therapy with simvastatin and bone marrow-derived mesenchymal stem cells increases benefits in infarcted swine hearts. *Arterioscler Thromb Vasc Biol*. 2009; 29: 2076-2083. doi: [10.1161/ATVBAHA.109.189662](https://doi.org/10.1161/ATVBAHA.109.189662)
20. Tang J, Wang J, Guo L, et al. Mesenchymal stem cells modified with stromal cell-derived factor 1 alpha improve cardiac remodeling via paracrine activation of hepatocyte growth factor in a rat model of myocardial infarction. *Mol Cells*. 2010; 29: 9-19. doi: [10.1007/s10059-010-0001-7](https://doi.org/10.1007/s10059-010-0001-7)
21. Gneocchi M, He H, Melo LG, et al. Early beneficial effects of bone marrow-derived mesenchymal stem cells overexpressing akt on cardiac metabolism after myocardial infarction. *Stem Cells*. 2009; 27: 971-979. doi: [10.1002/stem.12](https://doi.org/10.1002/stem.12)
22. Haider H, Jiang S, Idris NM, Ashraf M. Igf-1-overexpressing mesenchymal stem cells accelerate bone marrow stem cell mobilization via paracrine activation of sdf-1alpha/cxcr4 signaling to promote myocardial repair. *Circ Res*. 2008; 103: 1300-1308. doi: [10.1161/CIRCRESAHA.108.186742](https://doi.org/10.1161/CIRCRESAHA.108.186742)
23. Das R, Jahr H, van Osch GJ, Farrell E. The role of hypoxia in bone marrow-derived mesenchymal stem cells: Considerations for regenerative medicine approaches. *Tissue Eng Part B Rev*. 2010; 16: 159-168. doi: [10.1089/ten.teb.2009.0296](https://doi.org/10.1089/ten.teb.2009.0296)
24. Huang J, Zhang Z, Guo J, et al. Genetic modification of mesenchymal stem cells overexpressing ccr1 increases cell viability, migration, engraftment, and capillary density in the injured myocardium. *Circ Res*. 2010; 106: 1753-1762. doi: [10.1161/CIRCRESAHA.109.196030](https://doi.org/10.1161/CIRCRESAHA.109.196030)
25. Shujia J, Haider HK, Idris NM, Lu G, Ashraf M. Stable therapeutic effects of mesenchymal stem cell-based multiple gene delivery for cardiac repair. *Cardiovasc Res*. 2008; 77: 525-533. doi: [10.1093/cvr/cvm077](https://doi.org/10.1093/cvr/cvm077)
26. Tang YL, Zhao Q, Qin X, et al. Paracrine action enhances the effects of autologous mesenchymal stem cell transplantation on vascular regeneration in rat model of myocardial infarction. *Ann Thorac Surg*. 2005; 80: 229-236. doi: [10.1016/j.athoracsur.2005.02.072](https://doi.org/10.1016/j.athoracsur.2005.02.072)
27. Tang YL, Zhu W, Cheng M, et al. Hypoxic preconditioning enhances the benefit of cardiac progenitor cell therapy for treatment of myocardial infarction by inducing cxcr4 expression. *Circ Res*. 2009; 104: 1209-1216. doi: [10.1161/CIRCRESAHA.109.197723](https://doi.org/10.1161/CIRCRESAHA.109.197723)
28. Rosova I, Dao M, Capoccia B, Link D, Nolte JA. Hypoxic preconditioning results in increased motility and improved therapeutic potential of human mesenchymal stem cells. *Stem Cells*.

2008; 26(8): 2173-2182. doi: [10.1634/stemcells.2007-1104](https://doi.org/10.1634/stemcells.2007-1104)

29. Sanada F, Kim J, Czarna A, et al. C-kit-positive cardiac stem cells nested in hypoxic niches are activated by stem cell factor reversing the aging myopathy. *Circ Res*. 2014; 114: 41-55. doi: [10.1161/CIRCRESAHA.114.302500](https://doi.org/10.1161/CIRCRESAHA.114.302500)

30. Hahn JY, Cho HJ, Kang HJ, et al. Pre-treatment of mesenchymal stem cells with a combination of growth factors enhances gap junction formation, cytoprotective effect on cardiomyocytes, and therapeutic efficacy for myocardial infarction. *J Am Coll Card*. 2008; 51. doi: [10.1016/j.jacc.2007.11.040](https://doi.org/10.1016/j.jacc.2007.11.040)

31. Bartunek J, Behfar A, Dolatabadi D, et al. Cardiopoietic stem cell therapy in heart failure: The c-cure (cardiopoietic stem cell therapy in heart failure) multicenter randomized trial with lineage-specified biologics. *J Am Coll Cardiol*. 2013; 61: 2329-2338. doi: [10.1016/j.jacc.2013.02.071](https://doi.org/10.1016/j.jacc.2013.02.071)

32. Mastri M, Shah Z, McLaughlin T, et al. Activation of toll-like receptor 3 amplifies mesenchymal stem cell trophic factors and enhances therapeutic potency. *Am J Physiol Cell Physiol*. 2012; 303: C1021-C1033. doi: [10.1152/ajpcell.00191.2012](https://doi.org/10.1152/ajpcell.00191.2012)

33. Suzuki G, Weil BR, Young RF, Canty JM. Toll-like receptor 3 activation of cardiosphere-derived cells amplifies myocyte regeneration and reverse LV remodeling in swine with ischemic cardiomyopathy. *Circulation*. 2016; 134: A14423.

34. Hu S, Huang M, Nguyen PK, et al. Novel microRNA pro-survival cocktail for improving engraftment and function of cardiac progenitor cell transplantation. *Circulation*. 2011; 124: S27-S34. doi: [10.1161/CIRCULATIONAHA.111.017954](https://doi.org/10.1161/CIRCULATIONAHA.111.017954)

35. Dakhllallah D, Zhang J, Yu L, Marsh CB, Angelos MG, Khan M. MicroRNA-133a engineered mesenchymal stem cells augment cardiac function and cell survival in the infarct heart. *J Cardiovasc Pharmacol*. 2015; 65: 241-251. doi: [10.1097/FJC.0000000000000183](https://doi.org/10.1097/FJC.0000000000000183)

36. Gaetani R, Feyen DA, Verhage V, et al. Epicardial application of cardiac progenitor cells in a 3d-printed gelatin/hyaluronic acid patch preserves cardiac function after myocardial infarction. *Biomaterials*. 2015; 61: 339-348. doi: [10.1016/j.biomaterials.2015.05.005](https://doi.org/10.1016/j.biomaterials.2015.05.005)

37. Simpson D, Liu H, Fan TH, Nerem R, Dudley SC, Jr. A tissue engineering approach to progenitor cell delivery results in significant cell engraftment and improved myocardial remodeling. *Stem Cells*. 2007; 25: 2350-2357. doi: [10.1634/stemcells.2007-0132](https://doi.org/10.1634/stemcells.2007-0132)

38. Frederick JR, Fitzpatrick JR 3rd, McCormick RC, et al. Stromal cell-derived factor-1 α activation of tissue-engineered endothelial progenitor cell matrix enhances ventricular function after myocardial infarction by inducing neovasculogenesis.

Circulation. 2010; 122: S107-S117. doi: [10.1161/CIRCULATIONAHA.109.930404](https://doi.org/10.1161/CIRCULATIONAHA.109.930404)

39. Christman KL, Vardanian AJ, Fang Q, Sievers RE, Fok HH, Lee RJ. Injectable fibrin scaffold improves cell transplant survival, reduces infarct expansion, and induces neovasculature formation in ischemic myocardium. *J Am Coll Cardiol*. 2004; 44: 654-660. doi: [10.1016/j.jacc.2004.04.040](https://doi.org/10.1016/j.jacc.2004.04.040)

40. Liu J, Hu Q, Wang Z, et al. Autologous stem cell transplantation for myocardial repair. *Am J Physiol Heart Circ Physiol*. 2004; 287: H501-511. doi: [10.1152/ajpheart.00019.2004](https://doi.org/10.1152/ajpheart.00019.2004)

41. Xiong Q, Hill KL, Li Q, et al. A fibrin patch-based enhanced delivery of human embryonic stem cell-derived vascular cell transplantation in a porcine model of postinfarction left ventricular remodeling. *Stem Cells*. 2011; 29: 367-375. doi: [10.1002/stem.580](https://doi.org/10.1002/stem.580)

42. Gaetani R, Yin C, Srikumar N, et al. Cardiac-derived extracellular matrix enhances cardiogenic properties of human cardiac progenitor cells. *Cell Transplantation*. 2016; 25: 1653-1663. doi: [10.3727/096368915X689794](https://doi.org/10.3727/096368915X689794)

43. Lin YD, Yeh ML, Yang YJ, et al. Intramyocardial peptide nanofiber injection improves postinfarction ventricular remodeling and efficacy of bone marrow cell therapy in pigs. *Circulation*. 2010; 122: S132-S141. doi: [10.1161/CIRCULATIONAHA.110.939512](https://doi.org/10.1161/CIRCULATIONAHA.110.939512)

44. Danoviz ME, Nakamuta JS, Marques FL, et al. Rat adipose tissue-derived stem cells transplantation attenuates cardiac dysfunction post infarction and biopolymers enhance cell retention. *PLoS One*. 2010; 5: e12077. doi: [10.1371/journal.pone.0012077](https://doi.org/10.1371/journal.pone.0012077)

45. Itoh M, Nakayama K, Noguchi R, et al. Scaffold-free tubular tissues created by a bio-3d printer undergo remodeling and endothelialization when implanted in rat aortae. *PLoS One*. 2015; 10: 1-15. doi: [10.1371/journal.pone.0136681](https://doi.org/10.1371/journal.pone.0136681)

46. Laschke MW, Menger MD. Life is 3D: Boosting spheroid function for tissue engineering. *Trends in Biotechnology*. 2016. doi: [10.1016/j.tibtech.2016.08.004](https://doi.org/10.1016/j.tibtech.2016.08.004)

47. NI, Hibino N, Nakayama K. Principles of the kenzan method for robotic cell spheroid-based three-dimensional bioprinting. *Tissue Eng Part B Rev*. 2017; 23: 237-244. doi: [10.1089/ten.teb.2016.0322](https://doi.org/10.1089/ten.teb.2016.0322)

48. Williams AR, Hatzistergos KE, Addicott B, et al. Enhanced effect of combining human cardiac stem cells and bone marrow mesenchymal stem cells to reduce infarct size and to restore cardiac function after myocardial infarction. *Circulation*. 2013; 127: 213-223. doi: [10.1161/CIRCULATIONAHA.112.131110](https://doi.org/10.1161/CIRCULATIONAHA.112.131110)

49. Malliaras K, Smith RR, Kanazawa H, et al. Validation of

- contrast-enhanced magnetic resonance imaging to monitor regenerative efficacy after cell therapy in a porcine model of convalescent myocardial infarction. *Circulation*. 2013; 128: 2764-2775. doi: [10.1161/CIRCULATIONAHA.113.002863](https://doi.org/10.1161/CIRCULATIONAHA.113.002863)
50. Malliaras K, Li TS, Luthringer D, et al. Safety and efficacy of allogeneic cell therapy in infarcted rats transplanted with mismatched cardiosphere-derived cells. *Circulation*. 2012; 125: 100-112. doi: [10.1161/CIRCULATIONAHA.111.042598](https://doi.org/10.1161/CIRCULATIONAHA.111.042598)
51. Suncion VY, Ghersein E, Fishman JE, et al. Does transendocardial injection of mesenchymal stem cells improve myocardial function locally or globally?: An analysis from the percutaneous stem cell injection delivery effects on neomyogenesis (poseidon) randomized trial. *Circ Res*. 2014; 114: 1292-1301. doi: [10.1161/CIRCRESAHA.114.302854](https://doi.org/10.1161/CIRCRESAHA.114.302854)
52. Makkar RR, Smith RR, Cheng K, et al. Intracoronary cardiosphere-derived cells for heart regeneration after myocardial infarction (caduceus): A prospective, randomised phase 1 trial. *Lancet*. 2012; 379: 895-904. doi: [10.1016/S0140-6736\(12\)60195-0](https://doi.org/10.1016/S0140-6736(12)60195-0)
53. Malliaras K, Makkar RR, Smith RR, et al. Intracoronary cardiosphere-derived cells after myocardial infarction: Evidence for therapeutic regeneration in the final 1-year results of the caduceus trial. *J Am Coll Card*. 2014; 63: 110-120. doi: [10.1016/j.jacc.2013.08.724](https://doi.org/10.1016/j.jacc.2013.08.724)
54. Tokita Y, Tang XL, Li Q, et al. Repeated administrations of cardiac progenitor cells are markedly more effective than a single administration: A new paradigm in cell therapy. *Circ Res*. 2016; 119: 635-651. doi: [10.1161/CIRCRESAHA.116.308937](https://doi.org/10.1161/CIRCRESAHA.116.308937)
55. Gavira JJ, Nasarre E, Abizanda G, et al. Repeated implantation of skeletal myoblast in a swine model of chronic myocardial infarction. *Eur Heart J*. 2010; 31: 1013-1021. doi: [10.1093/eurheartj/ehp342](https://doi.org/10.1093/eurheartj/ehp342)
56. Malliaras K, Marban E. Cardiac cell therapy: Where we've been, where we are, and where we should be headed. *Br Med Bull*. 2011; 98: 161-185. doi: [10.1093/bmb/ldr018](https://doi.org/10.1093/bmb/ldr018)
57. Sanganalmath SK, Bolli R. Cell therapy for heart failure: A comprehensive overview of experimental and clinical studies, current challenges, and future directions. *Circ Res*. 2013; 113: 810-834. doi: [10.1161/CIRCRESAHA.113.300219](https://doi.org/10.1161/CIRCRESAHA.113.300219)
58. Tseliou E, Fouad J, Reich H, et al. Fibroblasts rendered antifibrotic, antiapoptotic, and angiogenic by priming with cardiosphere-derived extracellular membrane vesicles. *J Am Coll Cardiol*. 2015; 66(6): 599-611. doi: [10.1016/j.jacc.2015.05.068](https://doi.org/10.1016/j.jacc.2015.05.068)
59. Chakravarty T, Makkar RR, Ascheim DD, et al. ALLogenic heart STem cells to achieve myocardial regeneration (ALLSTAR) trial: Rationale and design. *Cell Transplant*. 2017; 26: 205-214.